Course Syllabus MCB 200A/B Fundamentals of Molecular and Cell Biology Fall Semester, 2013

Class Meetings: Monday-Friday 9:00-11:00 Location 125 LKS

Course Description: The goal of this course is to provide graduate-level instruction on molecular and cellular biosciences from a highly integrated systems perspective. A collection of approaches, and a focus on critical thinking and problem solving, will be used to show how fundamental, highly significant biological problems are "cracked open." Reading will be assigned from a mix of classic and current peer-reviewed papers selected by the instructors.

What this course aims to achieve:

- to provide exposure to the full breadth of research areas in Molecular and Cell Biology
- to provide enough depth to allow any graduate student to attend and understand any seminar in MCB
- to present material in a conceptual and integrated way that makes connections between apparently disparate research areas
- to demonstrate how an understanding of fundamental molecular and cell biological mechanisms is essential for understanding complex biological systems
- to interest students in research areas they may have not previously considered for their thesis work
- to cover material at the level of sophistication appropriate for a graduate level course, including a discussion of the experimental basis for what is being learned
- to equip students to approach the questions of 21st century biology
- to emphasize the value of basic research for addressing problems of human health and disease
- to emphasize the importance of clear and critical writing in biological research
- to teach students how to critically read and evaluate the primary literature
- to help prepare students for qualifying exams
- to ensure that students have the appropriate quantitative training to pursue cutting-edge research
- to help students become comfortable with asking questions during lectures and discussing science in a group setting

What this course does not aim to achieve:

- to provide a comprehensive coverage of every important topic within each area of MCB (this can be achieved in the Advanced Topics classes in the Spring)
- to provide remedial undergraduate-level training. Necessary background will be covered rapidly and it is expected that every student will have some areas where they will need to refresh or establish their mastery of the fundamentals.
- to provide a disconnected sequence of modules on each research area within MCB

Course Professors:

Dr. Don Rio (DR) Dr. David Drubin (DD) Dr. Marla Feller (MF) Dr. Susan Marqusee (SM) Dr. Jasper Rine (JR) Dr. Russell Vance (RV)

Grading will be based on attendance, worksheets, class participation, and presentations during the course.

WORKSHOP PAPERS – There are five workshops, one on a paper from each set of divisional QE papers. The first one (hosted by Prof. Rine) is: <u>Tong et al. 2001 Science 294:2364-2368</u>. "Systematic genetic analysis with ordered arrays of yeast deletion mutants." Everyone is expected to have read the paper before the coming to class. The other papers for later workshops will be announced in class.

SEPTEMBER CLASSICS – during the first month of class, students will be assigned to groups that will prepare and present a 20 min synopsis and critique of a classic paper in the field. Details on preparing the presentation will be discussed in class.

PROBLEM SETS/GRADING – problems and recommended reading will be assigned on a per lecture basis. Each Monday, a set of questions based on the previous week's lectures will be posted on bspace and assigned as a problem set, to be turned in the following Monday. Students are encouraged to discuss the problems with each other; however, it is assumed that what the students hand in will be the results of their own writing. Direct copying of answers is highly discouraged. Students will grade each other's problem sets on a rotating basis.

The following is based on material provided by the Campus to be distributed to students at the beginning of the academic year:

The student community at UC Berkeley has adopted the following Honor Code:

"As a member of the UC Berkeley community, I act with honesty, integrity, and respect for others." The hope and expectation is that you will adhere to this code.

Collaboration and Independence: Reviewing lecture and reading materials and studying for exams can be enjoyable and enriching things to do with fellow students. This is recommended. Students are encouraged to discuss homework problems with each other; however, it is assumed that what the students hand in will be the results of their own writing. Direct copying of answers is highly discouraged.

Cheating: A good lifetime strategy is always to act in such a way that no one would ever imagine that you would even consider cheating. Anyone caught cheating will also be reported to the University Center for Student Conduct.

Plagiarism: To copy text or ideas from another source without appropriate reference is plagiarism and will result in a failing grade for your assignment and usually further disciplinary action. For additional information on plagiarism and how to avoid it, see, for example: http://gsi.berkeley.edu/teachingguide/misconduct/prevent-plag.html

Academic Integrity and Ethics: Cheating on exams and plagiarism are two common examples of dishonest, unethical behavior. Honesty and integrity are of great importance in all facets of life. They help to build a sense of self-confidence, and are key to building trust within relationships, whether personal or professional. There is no tolerance for dishonesty in the academic world, for it undermines what we are dedicated to doing – furthering knowledge for the benefit of humanity.

Your experience as a student at UC Berkeley is hopefully fueled by passion for learning and replete with fulfilling activities. And we also appreciate that being a student may be stressful. There may be times when there is temptation to engage in some kind of cheating in order to improve a grade or otherwise advance your career. This could be as blatant as having someone else sit for you in an exam, or submitting a written assignment that has been copied from another source. And it could be as subtle as glancing at a fellow student's exam when you are unsure of an answer to a question and are looking for some confirmation. One might do any of these things and potentially not get caught. However, if you cheat, no matter how much you may have learned in this class, you have failed to learn perhaps the most important lesson of all.

MCB200A/B Lecture Schedule Fall 2013

Date		Lecturer	Торіс
Thursday	29-Aug	Jasper	Introduction to the Course; Origins of Life
Friday	30-Aug	Jasper/Don	Discovery of central dogma/Nucleic Acid chemistry
Monday	2-Sep	No class	Labor Day
Tuesday	3-Sep	Susan/Jasper	Tree of Life; Covalent chemistry of amino acids
Wednesday	4-Sep	Marla	Physics of microscopy
Thursday	5-Sep	David	Microscopy: molecular & cellular length scales
Friday	6-Sep	Jasper	Workshop: How to read a paper (Tong et al)
Monday	9-Sep	David	Properties of the cytoplasm +Sept Classic Group 1: Golgi (1906); Cajal (1906) Mentor: MF
Tuesday	10-Sep	David	Membrane properties +Sept Classic Group 2: Sturtevant (1913) Mentor: RV
Wednesday	11-Sep	David	Membrane protein topology and biogenesis +Sept Classic Group 3: Stanley (1935) Mentor: DR
Thursday	12-Sep	Susan	Workshop: Visualizing and Manipulating Structures +Sept Classic Group 4: Luria Delbruck (1943) Mentor: JR
Friday	13-Sep	John Kuriyan	Guest Lecture: Crystallography +Sept Classic Group 5: Avery & Hershey/Chase (1952) Mentor: JR
Monday	16-Sep	Russell	Differentiation, ES/IPS cells (applications, e.g., knockouts) +Sept Classic Group 6: Medawar (1953) Mentor: RV
Tuesday	17-Sep	Jasper	Specification of cell types, the yeast model +Sept Classic Group 7: Wat./Crick&Franklin&Wilkins(1953) Mentor: DR
Wednesday	18-Sep	Don	DNA replication: concepts and mechanisms +Sept Classic Group 8: Meselson/Stahl (1958) Mentor: JR
Thursday	19-Sep	Jasper	Mutation +Sept Classic Group 9: Kornberg (1958) Mentor: DR
Friday	20-Sep	Susan	Workshop: NSF proposal workshop and preparation +Sept Classic Group 1: Koshland (1958) Mentor: SM
Monday	23-Sep	Jasper	Meiosis +Sept Classic Group 2: Jamieson/Palade (1967) Mentor: DD
Tuesday	24-Sep	Jasper	Recombination +Sept Classic Group 3: Kandel (1970) Mentor: MF
Wednesday	25-Sep	Don	DNA repair pathways and Genome Editing +Sept Classic Group 4: Blobel (1975) Mentor: DD
Thursday	26-Sep	Susan	Molecules and how they work; driving forces +Sept Classic Group 5: Gurdon (1975) Mentor: RV
Friday	27-Sep	Marla	Workshop: Introduction to statistics +Sept Classic Group 6: Englander (1988) Mentor: SM
Monday	30-Sep	Marla	Resting/action potential +Sept Classic Group 7: Hartwell (1989) Mentor: DD
Tuesday	1-Oct	Susan	Molecules and how they work; driving forces +Sept Classic Group 8: Wickner (1994) Mentor: SM
Wednesday	2-Oct	Susan	Molecules and how they work; driving forces +Sept Classic Group 9: Julius (2002) Mentor: MF
Thursday	3-Oct	Marla	ion channels/poisson distributions
Friday	4-Oct	Marla	Workshop: Paper TBA
Monday	7-Oct	Russell	Innate immune recognition
Tuesday	8-Oct	Russell	Antibodies
Wednesday	9-Oct	Susan	Binding and measurement
Thursday	10-Oct	Don	Protein-Nucleic Acid interactions
Friday	11-Oct	Jamie Cate	Guest Lecture: Translation

Date		Lecturer	Торіс
Monday	14-Oct	David	Complex machines; microtubule dynamic instability/kinesin/dynein
Tuesday	15-Oct	David	Complex machines; the mitotic spindle: function and fidelity
Wednesday	16-Oct	Russell	Microbiota 1
Thursday	17-Oct	Russell	Microbiota 2
Friday	18-Oct	Russell	Workshop: Winter et al
Monday	21-Oct	Russell	Infection
Tuesday	22-Oct	Russell/Don	Ig recombination/biochemistry of RAG proteins
Wednesday	23-Oct	Jasper/Don	Epigenetics/Chromatin remodeling and modifications
Thursday	24-Oct	Susan/Jamie Fraser	Workshop: Computational sequence analysis
Friday	25-Oct	Marla	Workshop: Real world of science: grants, money
Monday	28-Oct	Don	Enzymology: purification, characterization, and mechanisms
Tuesday	29-Oct	Susan	Timescales of conformational changes 1
Wednesday	30-Oct	Susan	Timescales of conformational changes 2
Thursday	31-Oct	A. Martin	Molecular Machines/Allostery
Friday	1-Nov	Susan	Workshop: Agard alphalytic protease
Monday	4-Nov	Jasper	Genetic approach to transcription regulation
Tuesday	5-Nov	Don	Molecular mechanisms of transcription
Wednesday	6-Nov	Don	RNA processing and function
Thursday	7-Nov	David	Complex machines: principles of actin assembly/myosin activity
Friday	8-Nov	David	Complex machines: actin-based cell motility, Brownian ratchets
Monday	11-Nov	No class	Veteran's Day
Tuesday	12-Nov	Don	Transposition, retroviral integration, and site-specific recombination
Wednesday	13-Nov	Mike Levine	Development
Thursdsay	14-Nov	Marla	Development of the brain
Friday	15-Nov	Russell	Workshop: How to give a talk (Bonus: myth of scientific method?
Monday	18-Nov	David	Single cell analysis
Tuesday	19-Nov	Michael Rape	Guest lecture: Networks and signaling
Wednesday	20-Nov	Susan	Synthetic Biology
Thursday	21-Nov	Marla	Synapses
Friday	22-Nov	David	Workshop: Dynamic Instability
Monday	25-Nov	Marla	Learning as a cell signaling process
Tuesday	26-Nov	Marla	Pick a sensory system
Wednesday	27-Nov	TBD	TBD
Thur	28-Nov	No class	Thanksgiving
Friday	29-Nov	No class	Holiday
Monday	2-Dec	Don	Workshop: Paper
Tuesday	3-Dec	Iswar Hariharan	Guest Lecture: Cancer
Wednesday	4-Dec	Russell	Choose your own adventure
Thursday	5-Dec	Nipam Patel	Guest Lecture: Evo/Devo
Friday	6-Dec	Wrap session – and p	olan for a party in the evening (TBD)

Dec 9- STUDENT MICROSYMPOSIUM PRESENTATIONS (9-12pm) Dec 13