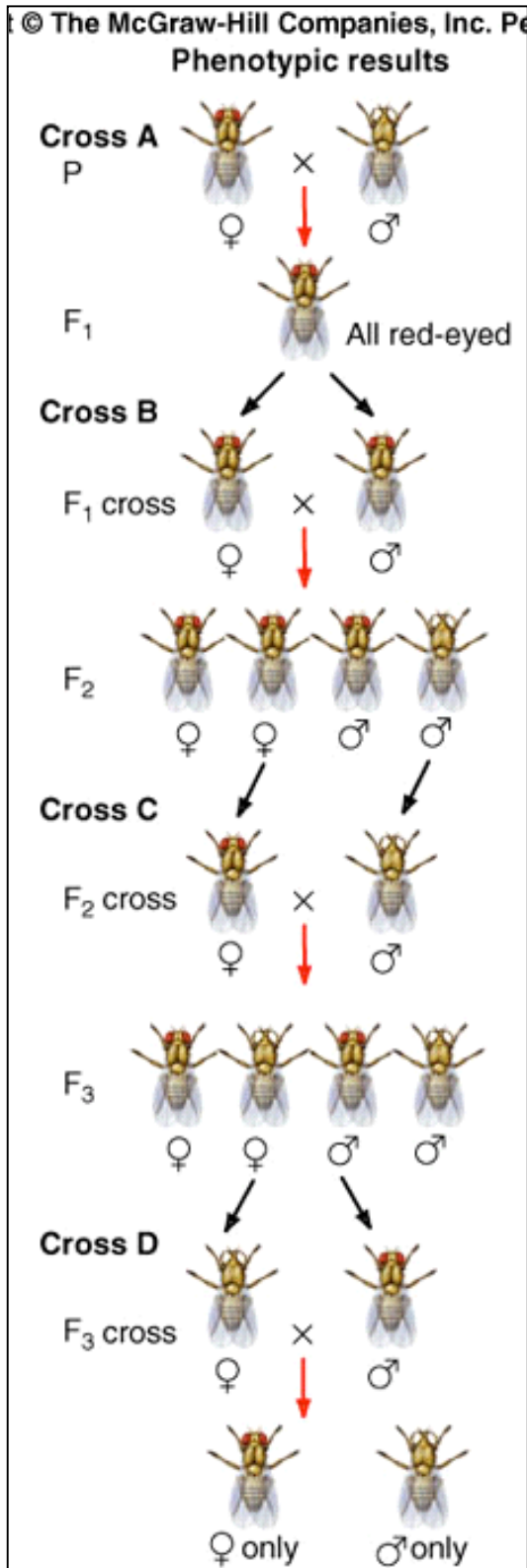


Question 2 (30 points)

A series of experiments by Thomas Hunt Morgan, and his student, Calvin Bridges, on eye color inheritance in *Drosophila*, form the centerpiece of the “chromosomal fact of inheritance.” A summary of crosses performed by Morgan is shown below.



This is hypothesis-driven science at its best. Write out the central hypothesis that T.H. Morgan pursued in his studies (one short sentence) – 10 points

Morgan hypothesized that the gene that specifies red eye color resides on the X chromosome

Answers such as “genes reside on chromosomes” get 5 points. Answers such as “eye color is sex-linked” get 7 points.

Now, pick one cross (just one) of all the ones shown on the left, write it out here (do not use genetic notation, just verbal descriptions of flies and their pedigrees), and explain, what specific consequence of his hypothesis Morgan was testing in this particular cross, and how the data from the cross supported the hypothesis. That part of the answer should take the form “if it is, in fact, true that ..., then one would expect that ...” (20 points)

A sample answer (several such answers are possible, and will be correct):

Cross C was between a red-eyed female, herself the daughter of a white-eyed male and a red-eyed female, and a white-eyed male. If Morgan’s hypothesis is true, then one expects half the males and half the females from this cross to be white-eyed.

Question 3 (30 points)

In class, we discussed the following 3-point testcross done by Alfred Sturtevant: two *Drosophila* were mated: a red-eyed fly that lacked a cross-vein on the wings and had snapped wing edges to a vermilion-eyed, normally veined fly with regular wings. All the progeny were wild type. These were testcrossed to a fly with vermilion eyes, no cross-vein and snapped wings. Sturtevant observed 1448 progeny in 8 phenotypic classes, as shown on the left. Sturtevant then used these data to map the genes, which are, of course, nicely linked.

$v \cdot cv^+ \cdot ct^+$	580
$v^+ \cdot cv \cdot ct$	592
$v \cdot cv \cdot ct^+$	45
$v^+ \cdot cv^+ \cdot ct$	40
$v \cdot cv \cdot ct$	89
$v^+ \cdot cv^+ \cdot ct^+$	94
$v \cdot cv^+ \cdot ct$	3
$v^+ \cdot cv \cdot ct^+$	5
	<hr/> 1448

Many other *Drosophila* species exist, and many are the object of investigation. You decide to take on genetic mapping in *Drosophila virilis*, which appears to be closely evolutionarily related to *D. melanogaster* – in fact, you find that the same recessive mutations (vermillion, no cross-vein, cut wing edges) exist in this species. You perform the same cross that Sturtevant did, but using *Drosophila virilis*: a red-eyed fly that lacked a cross-vein on the wings and had snapped wing edges to a vermilion-eyed, normally veined fly with regular wings. Not surprisingly, all the progeny are wild type. You testcross them to a fly with vermilion eyes, no cross-vein and snapped wings, and are absolutely stunned to discover the proverbial “something completely different.” In analyzing 1,000 progeny from this cross, you find the following animals in your vials:

Red eyes, no cross-vein, cut wing edges	395
Vermillion eyes, normal cross-vein, normal wings	405
Wild-type	98
Vermillion eyes, no cross-vein, normal wings	102

Once you’ve recovered from the emotional shock of having seemingly disproven a central tenet of genetics, you do two things (not necessarily in that order):

1. Come up with a hypothesis that explains the striking difference in the mapping data between *D. melanogaster* and *D. virilis* (10 points)

The ct and cv genes appear to have relocated very closely to each other in D. virilis (any form of statement that communicates the fact that the two genes are directly next to each other gets full credit)

2. Map – to the best of your ability – the genes in the *D. virilis* genome. Show your work. (20 points)

$v - 20 \text{ mu} - ct/cv$

(cannot determine order of the latter two from the data)

Question 4 (10 points)

You have two haploid yeast strains, one with a mutation at locus A and one with a mutation at locus B. The two loci are on different chromosomes. You mate the strains to form a diploid zygote, then sporulate (force the diploid to undergo meiosis) to form haploids. You randomly select 1000 of these haploid progeny, grow them up into cultures, and do a DNA-based experiment to test the genotypes. You are surprised to find that they do not obey Mendel's second law. Instead of seeing 25% of each possible genotype, you see that a third of the progeny are wild-type, a third have only the A mutation, and a third have only the B mutation. Propose a model to interpret these data which incorporates ONLY the A and B mutations and NO OTHER aberrations. Don't forget that the strains are haploid!

Answer: The combination of A and B is lethal. Any strain inheriting both mutations has died. The remainder of the genotypes appear at equal frequency in the progeny.

Question 5 (20 points)

George Beadle and Edward Tatum studied *Neurospora* strains carrying mutations that affected different stages of the arginine biosynthesis pathway. You get ahold of one of their Class II mutants, which can grow on minimal media supplemented with late precursor molecules (ornithine, citrulline, or arginine) but cannot grow on minimal media supplemented with precursor molecules which are used to make ornithine, *i.e.* which are upstream of ornithine in the pathway. In this strain, you introduce a mutation in the enzyme that converts citrulline to arginine. You then use some tricks, which are not important here, to mate your double mutant to the original Beadle and Tatum single mutant, creating a diploid. Assuming a model in which each mutation is equivalent to a deletion of the gene encoding an enzyme, what growth behavior do you expect from this diploid on:

- (a) Minimal media supplemented with precursors upstream of ornithine
- (b) Minimal media supplemented with citrulline.

Answer: (a), diploid cannot grow. (b), Diploid can grow.

Question 6 (30 points)

In class, we discussed four experiments that discovered conjugation in bacteria (Lederberg), its unidirectional nature (Hayes), the “spaghetti hypothesis” of the genome being threaded from one bacterium to another (Lwoff and Jacob/Monod), and phage-based transfer of genes from one bacterium to another, i.e., transduction (Zinder and Lederberg).

Consider a novel, pathogenic strain of *E. coli* that appears to be resistant to amoxicillin (a commonly used antibiotic that kills wild-type *E. coli*)

Describe, in a numbered list format, how you would ... (10 points each)

A. ... confirm that amo^R is an actual gene in the *E. coli* genome, i.e., that resistance to amoxicillin is genetic.

The L-D experiment in its direct form, if done on the resistant bacteria, is inapplicable – all cultures will yield a large number of resistant colonies. Three possible answers:

L-D on the sensitive bacteria

- 1. Take wt bacteria (start clonal and pool cultures)*
- 2. Expose to amo*
- 3. Count number of resistant colonies*
- 4. Compare*

Direct genetic transfer – via conjugation or transduction

- 1. Mate resistant and sensitive bacteria (or infect resistant with phage)*
- 2. Score for acquisition of resistance*

Amo “washout”

- 1. Culture resistant bacteria for an extended period of time in non- amo medium*
- 2. Expose to amo*
- 3. Number of resistant colonies should not drop significantly*

B. ... map the location of the amo^R gene on the *E. coli* chromosome relative to the *leu* genetic marker (the same one that Lederberg used). You are free to assume the existence of any bacterial strains or other reagents and biological materials you wish, but for your answer to be considered correct, you must explicitly state that you are assuming that they exist. (20 points)

Do an interrupted mating experiment with a $leu^+ amo^R$ bacterium and a $leu^- amo^S$ bacterium.

C. ... prove that the amo^R gene you have just identified and mapped is sufficient to endow resistance to amoxicillin to a strain of *E. coli* that, for unknown reasons, does not engage in conventional F-factor based conjugation, but appears to be a normal bacterium in every other respect.

Infect the amo^R -bacterium with lytic phage, take the phage supernatant, and infect the other bacterium with that phage, then plating those infected bacteria on amoxicillin under conditions that favor phage latency. Some students will propose a recombinant DNA-based way to do this, which is fine.